

## 2x Pre-Aliquoted Extensor Hi-Fidelity ReddyMix™ PCR Master Mix, 50µl Reaction

**Description:** Extensor Hi-Fidelity ReddyMix™ PCR Master Mix is a pre-aliquoted ready-to-use enzyme mix for long and accurate PCR without the need to mix individual components, reducing the risk of contamination and pipetting errors. The Extensor PCR Enzyme Mix, dNTPs, Extensor Reaction Buffer and MgCl<sub>2</sub> are all present in the mix. ReddyMix™ Master Mix also contains a dye and precipitant to facilitate gel loading.

**Ordering Information:** Select the buffer type from Table 1 and consumable type and size from Table 2.

Cat. No.	Buffer Type
EX1-xxx-8/LD	Buffer 1 - For templates <12kb
EX2-xxx-8/LD	Buffer 2 - For templates >12kb or problematic amplifications of any length.

**Note:** xxx denotes plastic consumable type

Cat. No.	Description	Quantity
EX?-266-8/LD/a	0.2ml Thermo-Strips & Domed Caps	50µl PCR x 12 strips of 8 tubes
EX?-266-8/LD/b	0.2ml Thermo-Strips & Domed Caps	50µl PCR x 10 x 12 strips of 8 tubes
EX?-337-8/LD/a	Individual 0.2ml Domed Cap Thermo-Tubes	50µl x 96 tubes
EX?-337-8/LD/b	Individual 0.2ml Domed Cap Thermo-Tubes	50µl x 10 x 96 tubes
EX?-350-8/LD/a	Individual 0.5ml Flat Cap Thermo-Tubes	50µl x 96 tubes
EX?-350-8/LD/b	Individual 0.5ml Flat Cap Thermo-Tubes	50µl x 10 x 96 tubes
EX?-489-8/LD/a	Individual 0.5ml Domed Cap Thermo-Tubes	50µl x 96 tubes
EX?-489-8/LD/b	Individual 0.5ml Domed Cap Thermo-Tubes	50µl x 10 x 96 tubes
EX?-600-8/LD	0.2ml Thermo-Fast® 96 Plates	50µl x 5 plates of 96 wells
EX?-620-8/LD/a	Individual 0.2ml Flat Cap Thermo-Tubes	50µl x 96 tubes
EX?-620-8/LD/b	Individual 0.2ml Flat Cap Thermo-Tubes	50µl x 10 x 96 tubes
EX?-700-8/LD	0.2ml Low Profile Thermo-Fast® 96 Plates	50µl x 5 plates of 96 wells
EX?-772-8/LD/a	0.2ml Low Profile Thermo-Strips & Domed Caps	50µl PCR x 12 strips of 8 tubes
EX?-772-8/LD/b	0.2ml Low Profile Thermo-Strips & Domed Caps	50µl PCR x 10 x 12 strips of 8 tubes
EX?-773-8/LD/a	0.2ml Low Profile Thermo-Strips & Flat Caps	50µl PCR x 12 strips of 8 tubes
EX?-773-8/LD/b	0.2ml Low Profile Thermo-Strips & Flat Caps	50µl PCR x 10 x 12 strips of 8 tubes
EX?-800-8/LD	0.2ml Skirted Thermo-Fast® 96 Plates	50µl x 5 plates of 96 wells
EX?-848-8/LD/a	0.2ml Low Profile Thermo-Strips & Domed Caps	50µl PCR x 8 strips of 12 tubes
EX?-848-8/LD/b	0.2ml Low Profile Thermo-Strips & Domed Caps	50µl PCR x 10 x 8 strips of 12 tubes
EX?-849-8/LD/a	0.2ml Low Profile Thermo-Strips & Flat Caps	50µl PCR x 8 strips of 12 tubes
EX?-849-8/LD/b	0.2ml Low Profile Thermo-Strips & Flat Caps	50µl PCR x 10 x 8 strips of 12 tubes

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**Table 2 (continued):**

Cat. No.	Description	Quantity
EX?-900-8/LD	0.2ml Semi Skirted Thermo-Fast® 96 Plates	50µl x 5 plates of 96 wells
EX?-990-8/LD	0.2ml Ultra Rigid Semi Skirted Thermo-Fast® 96 Plates	50µl x 5 plates of 96 wells
EX?-1000-8/LD	0.2ml Ultra Rigid Skirted Thermo-Fast® 96 Plates	50µl x 5 plates of 96 wells
EX?-1100-8/LD	0.2ml Thermo-Fast® 96 PCR Detection Plate	50µl x 5 plates of 96 wells
EX?-1300-8/LD	Thermo-Fast® 96 Robotic PCR Plate, Capped	50µl x 10 plates of 96 wells
EX?-1400-8/LD	Thermo-Fast® 96 PCR Detection Plate Mark II, Capped	50µl x 10 plates of 96 wells
EX?-1113-8/LD/a	0.2ml Thermo-Strips with Domed Caps	50µl PCR x 8 strips of 12 tubes
EX?-1113-8/LD/b	0.2ml Thermo-Strips with Domed Caps	50µl PCR x 10 x 8 strips of 12 tubes
EX?-1114-8/LD/a	0.2ml Thermo-Strips with Flat Caps	50µl PCR x 8 strips of 12 tubes
EX?-1114-8/LD/b	0.2ml Thermo-Strips with Flat Caps	50µl PCR x 10 x 8 strips of 12 tubes

**Note:** Replace ? with buffer type

**Example:** **EX1-337-8/LD/a**  
**2x Extensor Hi-Fidelity ReddyMix™ PCR Master Mix with Buffer 1, (Reaction Volume 50µl) Pre-aliquoted into 96 x 0.2ml Domed Cap Thermo-Tubes.**

**Kit Components:** Each well contains 25µl of a 2x working concentration PCR Master Mix. The addition of the template and primers (in a volume of 25µl) results in a final reaction volume of 50µl, containing:

**Buffer 1**

1x Extensor Buffer 1  
 1.25 units DNA Polymerase  
 2.25mM MgCl<sub>2</sub>  
 350µM each of dATP, dCTP,  
 dGTP and dTTP

**Buffer 2**

1x Extensor Buffer 2  
 1.25 units DNA Polymerase  
 2.25mM MgCl<sub>2</sub>  
 500µM each of dATP, dCTP,  
 dGTP and dTTP

**ReddyMix™ dye and precipitant**      **ReddyMix™ dye and precipitant**

Thermo-Fast® 96 plates are provided capped with Domed Cap Strips. An extra set of caps for application after the addition of template and primers is included with both plates and Thermo-Strips.

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**Overview of Protocol:**

For a 50µl reaction, take 25µl of Extensor Master Mix and add template, primers and water in a 25µl volume (scale up or down accordingly if required). Generally, 100–250ng template DNA, and 200nM (final concentration) of each primer is added. It is recommended that the Extensor Master Mix and added components are kept on ice. This removes the need for a hot start, as well as avoiding any degradation of primers and template through the 3' to 5' proofreading activity present in the Extensor Master Mix. The use of wax is not recommended, as it prevents adequate mixing of reaction components, leading to low yields. The enzyme mix has a fidelity that is at least four times higher than standard *Taq* DNA polymerase. All reaction tubes should be sterile and certified DNase/RNase free. The following points should also be noted:

- The Extensor Hi-Fidelity PCR Master Mix offers very robust amplification up to 15kb of human genomic DNA. Above 15kb, more optimisation may be required.
- Ensure proper mixing of reaction components, and always use thin-walled PCR tubes.
- Use a mineral oil overlay unless a heated lid thermocycler is used.
- Touchdown PCR may increase PCR product specificity.
- For best results, use primers (lengths 22–34 nucleotides) with annealing temperatures over 60°C.
- Primers can be used at 400nM for very long extensions.

**Templates:**

For the amplification of large DNA fragments, the quality of the template DNA is very important, as are the denaturation conditions. Keep template DNA denaturation steps as short as possible. Use Extensor Buffer 2 for DNA templates ≥ 12kb and when difficulties are expected or encountered. 125ng human genomic DNA is generally sufficient to provide good PCR results. When using simple templates (such as λ DNA), 1–10ng template DNA should prove sufficient; the number of cycles may be reduced by 5 and Extensor Buffer 1 can be used.

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### Thermal Cycler Programming:

For high fidelity PCR, a standard protocol should be used. For long PCR, modifications may need to be made. An example of a long PCR thermal cycling programme is given:

Initial denaturation	92-94°C <sup>1</sup>	2 min	1 cycle
Denaturation	92-94°C	10 sec	
Annealing <sup>2</sup>	50-68°C	30 sec	10 cycles
Extension	68°C <sup>3</sup>	x min <sup>4</sup>	
Denaturation	94°C	20 sec	
Annealing <sup>2</sup>	50-68°C	30 sec	15-20 cycles
Extension	68°C	x min (+20 s/cycle)	
Final extension	68°C	7 min	1 cycle

1 - When amplifying over 15kb, use a denaturation temperature of 92°C.

2 - Annealing temperature dependent on primers.

3 - Always use an extension temperature of 68°C, if possible. Often good results are obtained using a single annealing/extension step at 68°C.

4 - Extension times depend on the length of sequence to be amplified (see table below).

Amplicon size (kb)	3	6	10	20	30	40
Extension time (min)	2	4	8	15	20	30

After PCR, a sample (10–30% of reaction) may be loaded directly on a gel.

### Troubleshooting: **1 No product detected**

Try reducing the annealing temperature, increasing the concentration or quality of template, number of cycles or improving the purity of primers used.

### **2 Spurious bands appearing on electrophoresis gel**

When non-specific products are amplified, try increasing the annealing temperature (up to a maximum of 68°C) or reducing template concentration or cycle number.

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**Storage Conditions:** Can be stored at -20°C in a constant temperature freezer for up to 1 year. Avoid freeze thawing. Once opened, the vial can be stored at 4°C for up to 1 month. Shipped on ice.

**Tip:** The gel precipitant in ReddyMix™ Master Mix causes a slight increase in the thermal mass of the reaction mix. In a small number of cases this may necessitate some minor re-optimisation of the thermal cycler programme. If this is the case we suggest increasing the temperature of the denaturation step by 1–2°C and decreasing the temperature of the annealing step by 1–2°C. Alternatively, increase the duration of each step by 5–10 seconds.

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